

 <p><b>Proposal for Task Force Consideration at the ISSC 2017 Biennial Meeting</b></p>	<p>a. <input checked="" type="checkbox"/> Growing Area  b. <input type="checkbox"/> Harvesting/Handling/Distribution  c. <input type="checkbox"/> Administrative</p>
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Proposal Subject	Receptor Binding Assay (RBA) for Paralytic Shellfish Poisoning (PSP) Toxicity Determination
Specific NSSP Guide Reference	Section IV. Guidance Documents Chapter II. Growing Areas. 11 Approved NSSP Laboratory Tests
Text of Proposal/ Requested Action	<p>4. Approved Limited Use Methods for Marine Biotxin Testing</p> <p>This submission presents the ‘Receptor Binding Assay (RBA) for Paralytic Shellfish Poisoning (PSP) Toxicity Determination’ for consideration as an NSSP Approved Limited Use Method. The RBA is a competition-based assay that employs radiolabeled saxitoxin (3H-STX) to compete with PSP toxins present in standards/samples for binding sites on natural receptors in the assay. Following incubation with the receptors, unbound 3H-STX is removed and the remaining labeled toxin is measured with a scintillation counter. The amount of remaining 3H-STX is inversely proportional to standard/sample toxicity.</p> <p>The RBA offers a high-throughput, sensitive, and quantitative alternative to the mouse bioassay (MBA), which has been the long-standing reference method for PSP toxicity. Further, the RBA eliminates the use of live animals for detection of these toxins. While the RBA still uses receptors prepared from animals, the number of animals required for analysis is significantly reduced. Using native receptors as the analytical recognition elements for the assay allows for a composite measure of overall toxicity, as opposed to toxin concentrations measured by liquid chromatographic methods that require conversion factors of equivalent toxicity to calculate the overall toxicity.</p> <p>The RBA has undergone AOAC single- and multi-laboratory validation and is designated through AOAC as an Official Method of Analysis (OMA 2011.27). Results from those studies, and additional data, are included in this proposal submission for the RBA to be considered for approval as an NSSP Approved Limited Use Method for Marine Biotxin Testing.</p>
Public Health Significance	Paralytic shellfish poisoning intoxications result from the consumption of seafood (primarily bivalve molluscs) contaminated with neurotoxins known as paralytic shellfish toxins (PSTs). This suite of toxins binds to voltage-gated sodium channels and may result in paralysis if enough toxin is consumed. In extreme cases when respiratory support is not available to the patient, the intoxication may prove fatal. Since the toxins cannot be destroyed during cooking and there is no way to remove the toxins from seafood, the best control strategy is to ensure that contaminated product never reaches the market. To protect public health,

	<p>harvesting closures are implemented when toxicity exceeds the guidance level of 80 micrograms saxitoxin equivalents per 100 grams of shellfish tissue. As such, accurate analytical methods are needed to monitor shellfish toxicity for making decisions regarding opening and closing shellfish growing areas accordingly. Acceptance of the RBA as an NSSP Approved Limited Use Method for PSP toxicity determination would provide monitoring and management programs with an additional tool that can be used for monitoring toxin levels and making regulatory decisions. Not only does the RBA eliminate the need for live animals for PSP testing, it is also more sensitive than the MBA, thereby providing an early warning system for monitoring programs as toxin levels begin to rise.</p>
Cost Information	<p>The estimated cost for a full 96-well plate assay is ~\$95.00. Including standards and samples with triplicate measurements (as well as three dilutions per sample to ensure the unknown samples fall within linear range of assay), the cost per sample for quantitative results would be ~\$13.60. If running multiple plates or in screening mode, sample costs would be reduced. Further, the filter plates used in the RBA differ from ELISA plates in that all reagents are added to each well as needed rather than already being a component of the plate, making it more practical and cost-effective to analyze samples when there is less than a full plate.</p>
Action by 2013 Laboratory Methods and Quality Assurance Review Committee	<ol style="list-style-type: none"> <li>1. Recommended approval of this method as an alternative to the mouse bioassay for PSP in mussels.</li> <li>2. Recommended approval of this method for Limited Use for clams and scallops for the purpose of screening and precautionary closure for PSP.</li> <li>3. Recommended referral of this proposal to an appropriate committee as determined by the Conference Chairman to address this method in oysters.</li> <li>4. Recommended Executive Office sends a letter to submitter to request a checklist for evaluation of labs using this method with said checklist to be submitted within three (3) months.</li> </ol>
Action by 2013 Task Force I	<p>Recommended adoption of Laboratory Method Review and Quality Assurance Committee recommendation on Proposal 13-114.</p>
Action by 2013 General Assembly	<p>Adopted recommendation of 2013 Task Force I on Proposal 13-114.</p>
Action by FDA May 5, 2014	<p>Concurred with Conference action on Proposal 13-114.</p>
Action by 2015 Laboratory Methods Review Committee	<p>Recommended referral of Proposal 13-114 to an appropriate committee as determined by the Conference Chair until additional data for oyster matrix are received.</p>
Action by 2015 Task Force I	<p>Recommended adoption of Laboratory Methods Review Committee recommendation on Proposal 13-114.</p>
Action by 2015 General Assembly	<p>Adopted the recommendation of Task Force I on Proposal 13-114.</p>
Action by FDA January 11, 2016	<p>Concurred with Conference action on Proposal 13-114.</p>